

## Microbes and disturbance



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The microbiology of disturbed environments is a vigorous area of research<sup>1</sup> that has benefited from cultivation-independent molecular techniques. Our challenge now is to embrace the concept of 'microbial landscapes' in all its complexity and recognise the multidisciplinary nature of molecular and microbial ecology.

In conventional studies microbes in disturbed environments are overlooked if they cannot be cultured. Similarly, molecular approaches that involve gene selection and amplification using the polymerase chain reaction (PCR) have limited capacity to detect novel organisms if they have divergent gene sequences or occur at very low levels<sup>2</sup>. The literature concerned with extreme and disturbed environments abounds with reports of new microbes with novel gene arrangements and so it is highly likely that there will indeed be organisms overlooked by cultivation and PCR dependent techniques<sup>3-7</sup>. However, researchers are increasingly attempting to address this bottleneck<sup>8</sup> using techniques that bypass<sup>9</sup> or refine the PCR step<sup>10-11</sup>. The latter is particularly important because PCR is such a useful tool in microbial ecology. A set of protocols for analysing microbial diversity that brings together the most efficient diversity assessment tools<sup>7</sup> would address some concerns.

There is a strong link between naturally extreme and disturbed environments, and although this article focuses on human induced extremes there are many other examples of disturbance that have not been covered here<sup>1</sup>. Acid mine drainage (AMD) is a serious environmental issue (Figures 1 and 2) that can be a natural process leading to the production of sulfuric acid<sup>12</sup>. However, in mining the process can be accelerated and microbes contribute to acid formation<sup>13</sup>. For a long time it was believed that two bacterial species – *Acidithiobacillus ferrooxidans* and *A. thiooxidans* – were the main species associated with AMD<sup>6</sup>, but molecular techniques have shown that many microbes are involved<sup>3,6</sup>. Sulfate reducing bacteria (SRB) can reduce the acidity resulting from AMD and can react with metals to form insoluble metal sulfides that precipitate out of solution. As a result of this buffering and metal precipitation there is a growing

interest in the use of engineered anaerobic SRB to remove heavy metals. These bioreactors have been successful in the laboratory<sup>14-15</sup> and in the field<sup>16</sup>.

In environments contaminated with hydrocarbons, recovered molecular sequences are related to organisms that can degrade these compounds<sup>4</sup> (Streten-Joyce *et al*, unpublished data). Similarly, a striking feature of heavy metal introduction to the environment is the ensuing modification of microbial communities and their activities<sup>17</sup>. Bacteria show resistance to almost all toxic metal ions of environmental concern but there appears to be no general mechanism of metal resistance<sup>18-20</sup>. Although it is known that microbes can immobilise metals, evidence now suggests that they may themselves be subject to mobilisation<sup>21</sup>. This has implications for subsurface transport models and mechanisms of metal remobilisation, and provides a case for factoring microbes into these models.

There is mounting evidence that gene insertions under the control of integrons are a fundamental aspect of microbial



Figure 1. East Branch of the Finnis River, Northern Territory, Australia, looking towards Rum Jungle, a legacy uranium/copper mine site leaching acid. The white material on the left-hand bank are evaporites from the AMD, the cobbles in the creek are algal mats and the pH is around 3.8. Photo courtesy Jenny Brazier, Charles Darwin University and Environmental Research Institute of the Supervising Scientist (eriss) – Darwin Office.

Figure 2. Evaporites (salts containing a range of metals) resulting from AMD from the Rum Jungle mine site. Photo courtesy of Jenny Brazier, Charles Darwin University and eriss.



adaptation and evolution<sup>27</sup>. This also raises questions about the current reliance on organism identification using 16S rRNA genes, which may be evolving at different rates from genes involved in metabolism<sup>7,22</sup>. An analysis of a microbial community using only organism identification may be less informative than an analysis of community using genes and proteins directly associated with the disturbance. This leads to the somewhat vexed issue of ecological relevance and *in situ* monitoring using microbes – i.e. which genes and proteins should be targeted to satisfy the requirement for ecological relevance?

Microbes are widely used to assess ecological health because they provide a trophic link to higher organisms<sup>23,24</sup> and they respond quickly to the bio-available fraction of contaminants<sup>25</sup>. Bacteria have the highest surface area to volume ratio of any living organism, and only a thin membrane forms the boundary between them and their environment. Thus, bacteria are very sensitive to even slight environmental fluctuations and as such should be excellent early indicators of environmental degradation<sup>26</sup>. Microbial assays are commonly used as indicators of hydrocarbon<sup>27</sup> and heavy metal<sup>28</sup> contamination. Despite their obvious sensitivity to change, one of the biggest impediments to the routine use of bacteria is their significant temporal and spatial variation<sup>29</sup>. This variation may be more manageable if microbes are viewed at a community level and the focus shifts to functionally significant genes rather than tracking particular species.

Recent advances have provided us with the tools to catalogue genomes, proteins and metabolic pathways of the total microbiota in environmental samples. There is a growing awareness that most microbes are actually part of a complex and highly coordinated microbial 'city' or biofilm<sup>30</sup>. This concept of 'microbial landscapes' opens the door to a more rigorous approach to microbial community analysis through the application of classical ecological principles<sup>30</sup>. Added to this is our increasing capacity to study genes, proteins and metabolic pathways at the community level, and this provides an opportunity to learn about functional significance and mechanisms of tolerance or resistance. By understanding how microbes survive and function in disturbed environments, we should gain an appreciation of their ecological relevance and their capacity to act as early indicators of change.

## References

1. Satyanarayana T, Raghukumar C & Shivaji S. Extremophilic microbes: Diversity and perspectives. *Current Science* 2005; 89:78-90.
2. Baker BJ, Tyson GW, Webb RI *et al.* Lineages of acidophilic archaea revealed by community genomic analysis. *Science* 2006; 314:1933-5.
3. Baker BJ & Banfield JF. Microbial communities in acid mine drainage. *FEMS Microbiol Ecol* 2003; 44:139-52.
4. Carson B, Kastner M, Bartlett D *et al.* Implications of carbon flux from the Cascadia accretionary prism: results from long-term, in situ measurements at ODP Site 892B. *Mar Geol* 2003; 198:159-80.
5. Marchesi JR, Weightman AJ, Cragg BA *et al.* Methanogen and bacterial diversity and distribution in deep gas hydrate sediments from the Cascadia Margin as revealed by 16S rRNA molecular analysis. *FEMS Microbiol Ecol* 2001; 34:221-8.
6. Leduc D, Leduc LG & Ferroni GD. Quantification of bacterial populations indigenous to acidic drainage streams. *Water Air Soil Pollut* 2002; 135:1-21.
7. Nemergut DR, Martin AP & Schmidt SK. Integron diversity in heavy-metal-contaminated mine tailings and inferences about integron evolution. *App Environ Microbiol* 2004; 70:1160-8.
8. Kemp PF & Aller JY. Bacterial diversity in aquatic and other environments: what 16S rDNA libraries can tell us. *FEMS Microbiol Ecol* 2004; 47:161-77.
9. Tyson GW, Chapman J, Hugenholtz P *et al.* Community structure and metabolism through reconstruction of microbial genomes from the environment. *Nature* 2004; 428:37-43.
10. Barlaan EA, Sugimori M, Furukawa S *et al.* Profiling and monitoring of microbial populations by denaturing high-performance liquid chromatography. *J Microbiol Meth* 2005; 61:399-412.
11. Danovaro R, Luna GM, Dell'Anno A *et al.* Comparison of two fingerprinting techniques, terminal restriction fragment length polymorphism and automated ribosomal intergenic spacer analysis, for determination of bacterial diversity in aquatic environments. *Appl Environ Microbiol* 2006; 72:5982-9.
12. Harries J. Acid mine drainage in Australia: its extent and potential future liability. *Supervising Scientist* 1997; Report 125 Supervising Scientist, Canberra.
13. Edwards KJ, Bond PL, Gihring TM *et al.* An archaeal iron-oxidizing extreme acidophile important in acid mine drainage. *Science* 2000; 287:1796-9.
14. Jong T & Parry DL. Evaluation of the stability of arsenic immobilized by microbial sulfate reduction using TCLP extractions and long-term leaching techniques. *Chemosphere* 2005; 60:254-65.
15. Jong T & Parry DL. Removal of sulfate and heavy metals by sulfate reducing bacteria in short-term bench scale upflow anaerobic packed bed reactor runs. *Water Res* 2003; 37:3379-89.
16. Zaluski MH, Trudnowski J, Canty M *et al.* Status and Performance of Engineered SRB Reactors for Acid Mine Drainage Control. In: *Proceedings of The Sixth International In Situ and On-Site Bioremediation Symposium* 2001. San Diego, CA: Battelle Press.
17. Große C, Grass G, Anton A *et al.* Transcriptional organization of the *czc* heavy-metal homeostasis determinant from *Alcaligenes eutrophus*. *J Bacteriol* 1999; 181:2385-93.
18. Ji G & Silver S. Bacterial resistance mechanisms for heavy metals of environmental concern. *J Ind Microbiol* 1995; 14:61-75.
19. Silver S. Bacterial resistances to toxic metal ions *Gene* 1996; 179:9-19.
20. Silver S. Genes for all metals – a bacterial view of the periodic table – The 1996 Thom Award Lecture. *J Ind Microbiol Biotechnol* 1998; 20:1-12.
21. Boulton S, Hand VL & Vaughan DJ. Microbial controls on metal mobility under the low nutrient fluxes found throughout the subsurface. The 2006 *Science of the Total Environment*; 372:299-305.
22. Bourdineaud J-P, Baudrimont M, Gonzalez P *et al.* Challenging the model for induction of metallothionein gene expression. *Biochimie* 2006; 88:1787-92.
23. Hader DP, Kumar HD & Smith RC *et al.* Effects on aquatic ecosystems. *J Photochem Photobiol B: Biology* 1998; 46:53-68.
24. Denaro R, D'Auria G, Di Marco G *et al.* Assessing terminal restriction fragment length polymorphism suitability for the description of bacterial community structure and dynamics in hydrocarbon-polluted marine environments *Environ Microbiol* 2005; 7:78-87.
25. Hayat S, Ahmad I, Azam ZM *et al.* Effect of long-term application of oil refinery wastewater on soil health with special reference to microbiological characteristics. *Bioresource Technology* 2002; 84:159-63.
26. McArthur JV. Bacteria as biomonitors. In: Rader RB, Batzer DP, Wissinger SA (eds). *Bioassessment and Management of North American Freshwater Wetlands* 2001. New York, USA: John Wiley & Sons. 2001:249-61.
27. Dawson JJC, Godsiffe EJ, Thompson IP *et al.* Application of biological indicators to assess recovery of hydrocarbon impacted soils. *Soil Biol Biochem* 2007; 39:164-77.
28. Ryan RP, Ryan DJ & Dowling DN. Multiple metal resistant transferable phenotypes in bacteria as indicators of soil contamination with heavy metals. *J Soils Sediments* 2005; 5:95-100.
29. Merkley M, Rader RB, McArthur JV *et al.* Bacteria as bioindicators in wetlands: bioassessment in the Bonneville Basin of Utah, USA. *Wetlands* 2004; 24:600-7.
30. Battin TJ, Sloan WT & Kjelleberg S *et al.* Microbial landscapes: new paths to biofilm research. *Nature* 2007; 5:76-81.